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EFFECT OF TEMPERATURE & RELATIVE HUMIDITY ON SPORULATION AND SPORE GERMINATION ON ALTERNARIA SOLANI INCIDENCE OF EARLY BLIGHT OF TOMATO

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ABSTRACTTomato output is severely hampered by Alternaria leaf blight, which is brought on by Alternaria solani. At
JNKVV Jabalpur, an experiment was conducted in a laboratory experiment (2019) to determine the impact of
temperature and relative humidity on the sporulation of Alternaria solani, it was discovered that, after 72
hours of inoculation, a temperature of 25°C and a relative humidity of 90% were ideal for sporulation.
Additionally examined were the effects of temperature and relative humidity on sporulation of Alternaria
solani on a host in an *in vitro* condition. Alternaria solani sporulation was detected on the leaves that were
kept in a moist environment. The maximum sporulation on leaves was seen when the temperature reached
25°C and the relative humidity or spore germination. The maximum spore germination occurred after
72 hours of incubation at a temperature of 25°C and a relative humidity of 90%. It was followed by 30°C
temperature and 100 per cent relative humidity.

Key words: Tomato, Alternaria blight, sporulation, Spore germination.

Introduction

Tomato (*Lycopersicon esculentum* Mill) ranks as one of the most crucial vegetable crops globally, second only to potatoes in significance. Widely cultivated worldwide, it serves as a primary source of essential vitamins A, B and C. India holds the second position in both tomato cultivation area and production volume. In India, tomatoes are cultivated across approximately 8.85 lakh hectares, yielding a production of 19,696.0 metric tons with a productivity rate of 24.4 metric tons per hectare during the 2016-17 period. The prominent tomatogrowing states in India include Orissa, Bihar, Karnataka, Andhra Pradesh, Maharashtra, Madhya Pradesh and Tamil Nadu. In Madhya Pradesh specifically, tomato cultivation covers an area of 100.00 thousand hectares, producing 3,102.00 metric tons with a productivity rate of 31.02 metric tons per hectare (Anon, 2017).

Tomato plants are susceptible to various diseases from seedling emergence to harvest. Among these, early blight caused by *Alternaria solani* stands out as the most critical and limiting factor in tomato production (Datar and Mayee, 1981). This disease significantly impacts yield, causing considerable losses (Munde *et al.*, 2013). The incidence of early blight varies with location, reaching up to 70 percent in some instances (Munde *et al.*, 2013 and Pachori and Sharma, 2016). The influence of weather conditions on disease development has been observed by researchers worldwide at different times (Sangeetha and Siddaramaih, 2007; Devi and Chanu, 2012). As morphological characters and phylogenetic analysis, *Alternaria solani* bear large, long- beaked and noncatenated spores (Simmons, 2000). The mycelium consists of septa, branched, light brown hyphae which turned darker with age. The conidiophores are short, 50-90 μ m long and dark in colour. Conidia are 120-296 ×12-20 μ m in size, beaked, muriform, dark colour and born singly. Conidia contained 5-10 transverse 3 septa and 1-5 longitudinal septa (Singh, 1987). In view of the above facts the present study was undertaken and results are embodied here in.

Material and Methods

Effect of temperature and relative humidity on sporulation of *Alternaria solani* on Culture and Host

To find the ideal temperature and relative humidity for sporulation of *Alternaria solani* on the culture and host, an experiment was carried out *in vitro*.

Twenty ml of media were dispensed into each sterilized Petri dish. Using a sterilized cork borer, fivemillimeter discs of pure *Alternaria solani* culture was cultivated on PDA for seven days were excised from the culture's edge. Each Petri dish was then inoculated with one culture disc at the center and inverted. To investigate sporulation, these inoculated Petri plates were subjected to various temperatures and relative humidity levels ranging from 15, 20, 25, 30 and 35°C and 60, 70, 80, 90 and 100% over different incubation periods. Conidia enumeration was conducted using a hemocytometer.

The leaves were properly cleansed in sterile water after being carefully collected from a field with specific symptoms. The Petri plates with moist blotter, glass slides and glass rods that came with them were used to create a moist chamber where the leaves were stored. The leaves were incubated at various relative humidity levels and temperatures including 60, 70, 80, 90 and 100 percent and 15, 20, 25, 30 and 35°C, respectively. Following the procedure outlined by Buxton and Mellanby (1934), these relative humidity levels were maintained using concentrated H_2So_4 and distilled water in Table 1. To record sporulation, one ml of distilled water was used to wash the leaves. Using a hemocytometer, sporulation was detected at various intervals. For each treatment, three replications were kept. Petri plates were checked for sporulation at 24, 48 and 72 hours. The measurement of sporulation was done with the aid of a hemocytometer.

Table 1: Quantity of H_2So_4 and distilled water for different
humidity level.

Humidity level (%)	$H_2So_4(ml)$	Distilled Water (ml)
60	37.5	62.5
70	32.5	67.5
80	26	74
90	10	90
100	0	100

 Table 2:
 Effect of temperature on sporulation of Alternaria solani on culture and host (In vitro).

	Incubation hours									
Temp.		Culture				Host				
(°C)	(Spo	s per	cm²)	(Spores per cm ²)						
	24		48	72	24		48	72		
15	96.96	1	06.7	138.88	295.92	3	04.48	326.7		
20	141.18	15	54.29	185.29	325.92	3	52.77	362.03		
25	190.55	21	18.89	256.55	365.73	3	67.59	412.03		
30	163.18	1	80.7	232.59	326.84	3	49.07	370.37		
35	147.59	16	52.33	197.03	306.47	3	10.44	319.44		
Factors	SEm:	±	CD	at 5%	SEm±	:	CD	at 5%		
Temp.(A)	8.32		24	4.14	12.23		1	N/A		
Hrs. (B)	6.44		N	J/A	9.48		1	N/A		
Interaction	14.41		41.82		21.19		61.52			
(A×B)	14.41		4	1.02	21.19		C	01.52		
*Average of Three replications.										

Effect of temperature and relative humidity on spore germination of *Alternaria solani* on culture

The goal of the experiment was to determine the ideal temperature and relative humidity for spore germination of *Alternaria solani*. For this, the cavity slide approach was used. Conidial suspension was created in culture tubes using sterile water. Each slide had two chambers, each of which received one ml of conidial suspension. For each treatment, three replications were retained. In a wet chamber created with Petri plates filled with a moist blotter and glass rods, cavity slides were stored. At 24, 48 and 72 hours, spore germination was observed.

In order to determine the impact of temperature and relative humidity on spore germination, cavity slides containing spore suspension were kept in Petri dishes at various temperatures and five relative humidity levels, including 15, 20, 25, 30 and 35° C and 60, 70, 80, 90 and 100% at various time intervals. These humidity levels were maintained by utilizing distilled water and H₂So₄ in various concentrations.

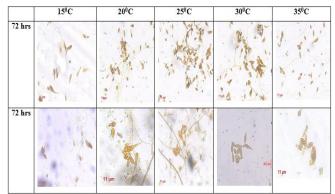


Fig. 1: Effect on temperature on sporulation of *Alternaria solani* on culture and host after 72 hrs of incubation.

Results and Discussion

Effect of temperature on sporulation of *Alternaria* solani on culture and host

The results shown in Table 2 and Fig. 1 in culture showed that temperature and incubation time had a substantial impact on Alternaria solani sporulation. After 72 hours of incubation, the highest level of sporulation was noted. Temperatures of 25, 30, 35, 20 and 15°C each had 232.59, 197.03, 185.29 and 138.88 spores per cm² as their maximum sporulation, respectively. Temperature and incubation time interacted significantly to affect sporulation. The maximum sporulation was recorded at 25°C temperature. Sharma and Ahir (2018), Somappa et al., (2013) and Aruna kumara (2006) have reported the similar observation that the sporulation of Alternaria solani at 25°C temperature followed by 30°C and 35°C temperature sporulation was recorded at 10°C temperature. The variations in sporulation of Alternaria solani in present studies, support the views of earlier workers (Ansari et al., 1989) who have reported that growth and sporulation was influenced by temperature.

In the host, the data shown in Table 2 and Fig. 1 showed that after 72 hours of incubation, the greatest sporulation of 412.03 per cm² was seen at a temperature of 25°C followed by 370.37 and 362.03 at 30 and 20°C respectively. At a temperature of 35°C *Alternaria solani* showed the least amount of sporulation. Sporulation was significantly impacted by the incubation time. The 25 and 30°C temperature also support the sporulation of *Alternaria solani*. The present results are coincided with the findings of Khare (1979) and also same result was found by Kaul and Saxena (1988).

Effect of relative humidity on sporulation of *Alternaria solani* on culture and host

The data in Table 3 and Fig. 2 showed that, in culture, sporulation varied with varying humidity and incubation

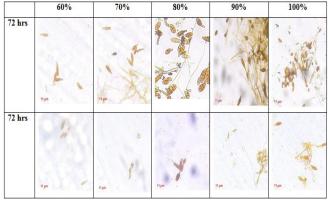


Fig. 2: Effect on relative humidity on sporulation of *Alternaria solani* on culture and host after 72 hrs of incubation.

 Table 3:
 Effect of relative humidity on sporulation of Alternaria solani on culture and host (In vitro).

Dala diana	Incubation hours								
Relative Humidity	Culture				Host				
	(Spores per cm ²)				(Spores per cm ²)				
(%)	24		48	72	24		48	72	
15	141.18	16	66.85	210.63	298.96	3	06.7	315.77	
20	163.55	201.18		244.66	321.84	3.	31.59	340.85	
25	198.51	229.51		300.74	330.81	34	40.29	363.4	
30	243.7	292.92		379.63	373.59	3	98.26	426.99	
35	208.7	25	54.73	332.48	336.10	3	71.03	400.44	
Factors	SEm± CD at 5%				SEm±	:	CD	at 5%	
Temp.(A)	8.903	;	25.839		10.435		30.283		
Hrs. (B)	6.897	'	N	J/A	8.083		1	N/A	
Interaction	15.421		44,755		18.073		52.452		
(A×B)	15.42	44		133	18.073		52	2.432	
*Average of Three replications.									

times. Both humidity and the length of the incubation time had a big impact on sporulation. At 90% relative humidity, the highest sporulation of 379.63 per cm² was reported. This was followed by 332.48, 300.74, 244.66 and 210.63 per cm² at 100%, 80%, 70% and 60% relative humidity respectively. The incubation period also had a big impact on sporulation. 72 hours of incubation with 90% relative humidity resulted in the highest sporulation, which was measured at 379.63 per cm². The maximum sporulation was recorded at 90 per cent relative humidity. Sharma and Ahir (2018), Somappa et al., (2013) and Aruna kumara (2006) have reported the similar observation that the maximum sporulation of Alternaria solani 90 per cent relative humidity followed by 100 per cent relative humidity and the minimum sporulation was recorded at 10°C temperature.

The data in Table 3 and Fig.2 make it clear that the influence of relative humidity and incubation period on sporulation in the host was determined to be statistically significant. Maximum sporulation occurred at a relative humidity of 90% reaching 426.99 spores per cm² with

	15°C	20 ⁰ C	25°C	30°C	35°C	
72 hrs	·/ ···································		1		1	
	60%	70%	80%	90%	100%	
72 hrs				1	A.	

Fig. 3: Effect on temperature and relative humidity on spore germination of *Alternaria solani* after 72 hrs of incubation.

Temp.	Incubation hours						
(°C)	24	%	48	24	%	48	
15	9.667 34.52		11.333	40.46	13.333	47.60	
20	13.000	46.42	16.667	59.53	18.000	64.28	
25	17.333	61.89	22.667	80.92	26.667	95.21	
30	14.333	51.17	18.667	66.64	19.333	69.03	
35	13.000	46.42	15.333	54.75	17.667	63.07	
Factors		SEm±		0	CD at 5%	, D	
Temp. (A)		0.725			2.105		
Hrs. (B)		0.562		1.556			
Interaction		1.256		3.645			
(A×B)		1.230		3.043			
*Average of Three replications.							

 Table 4:
 Effect of temperature on spore germination of Alternaria solani.

100, 80, 70 and 60 percent having 400.44, 363.4, 340.85 and 315.77 spores per cm² respectively. Statistically significant differences between incubation periods and sporulation were discovered. The relative humidity played vital role with references to sporulation. The relative humidity ranged from 60 to 100 per cent respectively. Maximum sporulation observed at 90 per cent relative humidity. The 90 and 100 per cent relative humidity also support the sporulation of *Alternaria solani*. The present results are coincided with the findings of Khare (1979).

Effect of temperature and relative humidity on spore germination of *Alternaria solani*

The information in Table 4 and Fig. 3 showed that temperature and incubation time had an impact on the spore germination. Temperature and incubation time had a substantial impact on the percentage of spore germination. After 72 hours of incubation, the temperature at which the maximum spore germination (95.21%) was observed was 25°C followed by temperatures of 69.03, 64.28, 63.07 and 47.60°C and temperatures of 20, 30 and 35°C respectively. Comparisons between various incubation periods were discovered to be statistically significant. The data shown in Table 5 and Fig. 3 showed that after 72 hours of incubation, the maximum spore germination (98.81%) was obtained at 90% relative humidity, followed by 84.52, 75.00, 61.89 and 55.92% at 100%, 80.7% and 60% relative humidity, respectively. Incubation time also had a substantial impact on the percentage of spores that germinated, and variations between them were also found to be statistically significant.

Five temperature (15, 20, 25, 30 and 35° C) and relative humidity (60, 70, 80, 90 and 100%) were used to observe their effect on spore germination of *A. solani*. Spore germination of *A. solani* varied with different

Table 5: Effect of relative humidity on spore germination ofAlternaria solani.

Relative	Incubation hours							
Humidity(%)	24	%	48	24	%	48		
60	12.000	42.85	13.333	47.60	15.667	55.92		
70	12.667	45.21	14.667	52.35	17.333	61.89		
80	14.667	52.35	17.667	63.07	21.000	75.00		
90	18.667	66.64	22.333	79.75	27.667	98.81		
100	17.000	60.71	20.333	72.60	23.667	84.52		
Factors		SEm± CD at 5%						
Temp.(A)		0.650	1.886					
Hrs. (B)		0.503		1.395				
Interaction		1.125		3.266				
(A×B)		1.125			3.200			
*Average of Three replications.								

temperature and relative humidity levels. Maximum spore germination was recorded in 25°C temperature and 90 per cent relative humidity against minimum spore germination at 15°C temperature and 60 per cent relative humidity. However, temperature 30°C and 100 per cent relative humidity had significant effect on spore germination of *A. solani*. Similar results were also reported by Stevenson and Pennypacker (1988).

Conclusion

It was concluded the conditions that favored the highest sporulation of *Alternaria solani* on culture were observed at 25°C with a relative humidity of 90% followed by 30°C with 100% humidity. Additionally, peak sporulation on the host leaf was observed at 25°C with a relative humidity of 90%. Conversely, the lowest sporulation was recorded at 15°C with a relative humidity of 60%. Optimal spore germination occurred at 25°C with a relative humidity of 90%, with secondary favorable conditions noted at temperatures of 30°C, 20°C, 35°C and 15°C along with relative humidity levels of 100%, 80%, 70% and 60%, respectively.

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Conflict of interest: On behalf of all authors, the corresponding author states that there is no conflict of interest exists.

References

Anonymous (2017). Horticultural Statistics at a Glance. National Horticulture Board, Department of Agriculture Cooperation, *Ministry of Agriculture*. 481.

Ansari, N.A., Khan M.W. and Muheet A. (1989). Effect of

some factors on growth and sporulation of *Alternaria* brassicae causing Alternaria blight of rapeseed and mustard. Acta Botanical Indica. **17**, 49-53.

- Aruna, kumara, K.T. (2006). Studies on Alternaria solani (Ellis and Martin) Jones and Grout causing early blight of tomato. Thesis, University of Agrilcultural Science, Dharwad. 70.
- Buxton, P.A. and Mellanby K. (1934). Measurement and control of humidity, *Bulletin of Entomology Res.* 25, 171-175.
- Datar, V.V. and Mayee C.D. (1981). Assessment of losses in tomato yield due to early blight. *Indian Phytopathol.* 34(2), 191-195.
- Devi, A.P. and Chanu L.B. (2012). Airspora and epidemiology of early blight of tomato caused by *Alternaria solani* (Ellis and Mart) Jones and Grout in Manipur. J. of Mycopathol. Res. 50(1), 81-84.
- Kaul, A.K. and Saxena H.K. (1988). Physiologic specialization in *Alternaria solani* causing early blight of tomato. *Indian J. Myco. Plant Patho.* 18, 128-132.
- Khare, U.K. (1979). Epidemiology and histopathology of Alternaria blight (*Alternaria porri* (Ellis) Ciferri) of onion (*Allium cepa*). Thesis, College of Agriculture, Jawaharlal Nehru Krishi Vishwa Vidyalaya, Jabalpur 66.

- Munde, V.G., Diwakar M.P., Thombre B.B. and Dey U. (2013). Survey and surveillance of early blight of tomato caused by *Alternaria solani* in Konkan region. *Int. J. of Plant Protection.* 6(2), 476-477.
- Pachori, A. and Sharma O.P. (2016). Status of early blight of tomato in morena, bhind and Gwalior districts of MP. *The Bioscan.* **11(2)**, 791-793.
- Sangeetha, C.G and Siddaramaiah A.L. (2007). Epidemiological studies of white rust, downy mildew and Alternaria blight of Indian mustard (*Brassica juncea* (Linn.) Czern. and Coss.). *African J. of Agril Res.* **2**(7), 305-308.
- Sharma, R.L. and Ahir R.R. (2018). Physiological studies of Alternaria solani causing Alternaria blight of tomato. J. of Entomology and Zoology studies. 6(6), 844-847.
- Simmons, E.G. (2000). *Alternaria* themes and variations (244-286) species on Solanaceae. *Mycotaxon.* **75**, 115.
- Singh, R.S. (1987). Diseases of Vegetable Crops. Oxford and IBH Publication *Pvt. Ltd. New Delhi.* 419.
- Somappa, J., Srivastava K., Sarma B.K., Pal C. and Kumar R. (2013). Studies on growth conditions of the tomato Alternaria leaf spot causing *Alternaria Solani* L. *The Bioscan.* **8(1)**, 101-104.
- Stevenson, R.E. and Packer S.P. (1988). Effect of radiation temperature and moisture on conidial germination of *Alternaria solani*. *Phytopathology*. **78**, 926-930.